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# Carbonic anhydrase inhibitors. Inhibition of the human cytosolic isozyme VII with aromatic and heterocyclic sulfonamides

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Abstract—The inhibition of a newly cloned human carbonic anhydrase (CA, EC 4.2.1.1), isozyme VII (hCA VII), has been investigated with a series of aromatic and heterocyclic sulfonamides, including some of the clinically used derivatives (acetazolamide, methazolamide, ethoxzolamide, dichlorophenamide, dorzolamide, brinzolamide and benzolamide), as well as the sulfamate antiepileptic drug topiramate. Inhibition data for the the other physiologically relevant cytosolic isoforms hCA I, hCA II and mCA XIII are also provided for comparison. hCA VII shows a high catalytic activity for the CO<sub>2</sub> hydration reaction, with a  $k_{\rm cat}$  of  $9.5 \times 10^5$  s<sup>-1</sup> and  $k_{\rm cat}/K_{\rm m}$  of  $8.3 \times 10^7$  M<sup>-1</sup> s<sup>-1</sup> at pH 7.5 and 20 °C. A very interesting inhibition profile against hCA VII with this series of 32 sulfonamides/sulfamates was observed. hCA VII shows high affinity for all the investigated compounds, with inhibition constants in the range of 0.45-210 nM. Topiramate, ethoxzolamide and benzolamide showed subnanomolar hCA VII inhibitory activity, whereas acetazolamide, methazolamide, dorzolamide and brinzolamide showed K<sub>I</sub>-s in the range of 2.1-3.5 nM. Dichlorophenamide was slightly less active (K<sub>I</sub> of 26.5 nM). A number of heterocyclic or bicyclic aromatic sulfonamides also showed excellent hCA VII inhibitory properties ( $K_{I}$ -s in the range of 4.3–7.0 nM) whereas many monosubstituted or disubstituted benzenesulfonamides were less active (K<sub>I</sub>-s in the range of 45–89 nM). The least active hCA VII inhibitors were some substituted benzene-1,3disulfonamides as well as some halogenated sulfanilamides ( $K_{\rm I}$ -s in the range of 100–210 nM). The inhibition profile of hCA VII is rather different of that of the other cytosolic isozymes, providing thus a possibility for the design of more selective, hCA VII-specific inhibitors. In addition, these data furnish further evidence that hCA VII is the isozyme responsible for the anticonvulsant/antiepileptic activity of sulfonamides and sulfamates. © 2004 Elsevier Ltd. All rights reserved.

## 1. Introduction

Vertebrates, humans included, express a multitude of isozymes of the metalloenzyme carbonic anhydrase (CA, EC 4.2.1.1), which play a host of physiological roles in processes where the three chemical entities acting as substrates/reaction products of the CAs (i.e., CO<sub>2</sub>, bicarbonate and H<sup>+</sup> ions) are involved. Such processes include respiration and gas exchange, pH homeostasis, renal excretion of anions and urine formation, detoxification of metabolic waste products, production of electrolytes in many tissues/organs (aqueous humor

in the eye, cerebrospinal fluid (CSF) in the brain, bicarbonate secretion in the kidney, pancreas and other organs, HCl production in the stomach, etc.), biosynthetic reactions, bone resorption, cell proliferation and differentiation, cell adhesion, modulation of neuronal transmission, etc. 1-6 In humans, 15 isozymes are presently known, twelve of which are catalytically active (CAs I-Va, CA Vb, CAVI, CA VII, CA IX and CAs XII-XIV), whereas the CA-related proteins (CARPs VIII, X and XI) are devoid of catalytic activity due to the fact that one or more histidine zinc ligand are replaced by other amino acid residues.<sup>7</sup> Among the active isozymes, five show a cytosolic subcellular localization: CAs I-III, CA VII and CA XIII. 1-5,8 Whereas CA III is a very low activity enzyme, abundant in the skeletal muscles and adipocytes (also poorly inhibited by

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**Table 1.** Kinetic parameters for  $CO_2$  hydration reaction catalyzed by the cytosolic  $\alpha$ -CA isozymes I–III, VII and XIII, at pH 7.5, and their inhibition data with acetazolamide **AAZ** (5-acetamido-1,3,4-thiadiazole-2-sulfonamide), a clinically used compound

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Isozyme	Activity level	$k_{\text{cat}}$ (s <sup>-1</sup> )	$k_{\text{cat}}/K_{\text{m}}$ $(M^{-1} \text{ s}^{-1})$	K <sub>I</sub> (acetazolamide) (nM)
hCA I*	Moderate	$2 \times 10^{5}$	$5 \times 10^7$	250
hCA II*	Very high	$1.4 \times 10^{6}$	$1.5 \times 10^{8}$	12
hCA III**	Low		$3 \times 10^{5}$	300,000
mCA VII**	High	$9.4 \times 10^{5}$	$7.6 \times 10^{7}$	16 <sup>*</sup>
mCA XIII*	Moderate	$8.3 \times 10^{4}$	$4.3 \times 10^{7}$	17

h = human; m = murine isozyme.

sulfonamides),<sup>1–5</sup> the other four are quite active (Table 1). Some isozymes (i.e., CA II) were known for decades to be physiologically important ones,<sup>1–5</sup> others (such as CA I) have physiological functions little understood at the moment,<sup>1–5</sup> whereas for CA XIII, an important role in pH regulation in the reproductive tract of both females and males has only recently been recognized.<sup>8</sup>

CA VII appears to be the less studied and understood among the cytosolic CAs. The CA VII was isolated from a human genomic library in 1991 by Montgomery et al., and found to be located on chromosome 16. The gene is approximately 10 kb long and contains seven exons and six introns found at positions identical to those determined for the previously identified CA I, CA II and CA III genes. The derived amino acid sequence of the CA VII gene is of 263 amino acids showing 50%, 56%, and 49% identity with hCA I, hCA II and hCA III, respectively (these were the cytosolic isozymes known in 1991). An initial purification of the mouse isozyme, mCA VII, by Tashian's group led to the conclusion that this is a low activity CA, by the conclusion that this is a low activity CA, by the conclusion that this is a low activity CA, by the conclusion that this is a low activity CA, by the conclusion that this is a low activity CA, by the conclusion that this is a low activity CA, by the conclusion that the conclus

work of the same group,<sup>11</sup> with a better purification protocol led to the thorough purification and characterization of mCA VII (in fact a chimeric protein, with the first 9 amino acids of human origin, and the remaining 253 of murine origin). This was found to be catalytically highly active ( $k_{\rm cat}/K_{\rm m}$  for CO<sub>2</sub> hydration described by two ionizations at p $K_{\rm a}$  6.2 and 7.5, with a maximum approaching  $8 \times 10^7 \, {\rm M}^{-1} \, {\rm s}^{-1}$ ) and inhibitable by sulfonamides such as the clinically used acetazolamide and ethoxzolamide (in the low nanomolar range—Table 1).<sup>11</sup> From these data it is apparent that CA VII is strikingly similar to the physiologically most relevant isozyme, CA II, which shows a comparable high  $k_{\rm cat}/K_{\rm m}$  for CO<sub>2</sub> hydration of  $1.5 \times 10^8 \, {\rm M}^{-1} \, {\rm s}^{-1}$ , and is also very sensitive to sulfonamide CA inhibitors (CAIs)—Table 1.<sup>1–5</sup>

From the data in Table 1, it is obvious that CA VII is the second most active cytosolic isozyme after CA II (showing approximately 70% of the hCA II activity), and it seems to be susceptible to inhibition by the clinically used sulfonamide CAI (although only two such derivatives have been investigated with the murine chimeric isozyme). 11 CA VII has been shown to be highly expressed in the brain, by in situ hybridization. 12,13 Very recently, one of our groups<sup>13</sup> showed that the capability to generate stimulation-induced synchronous action potential discharges emerges abruptly in the rat hippocampus at approximately postnatal day 12. This was directly attributable to a steep developmental up-regulation of CA VII in pyramidal neurons. 13 These results pointed to a crucial role for the developmental expression of the isozyme VII activity in shaping long-term plasticity and promoting epileptogenesis.

The human isozyme hCA VII has not been cloned and characterized. Here we present such a study, as well as the first investigation of sulfonamide inhibition with this

<sup>\*</sup>From Ref. 8a.

<sup>\*\*</sup> From Ref. 11.

isozyme. Sulfonamides, such as, acetazolamide **AAZ**, methazolamide **MZA**, ethoxzolamide **EZA** and dichlorophenamide **DCP**, have been clinically used for more than 40 years as systemic CAIs, whereas two additional drugs dorzolamide **DZA** and the structurally-related brinzolamide **BRZ** are topically acting antiglaucoma CAIs used since 1995. <sup>1–5</sup> Topiramate, **TPM**, an antiepileptic with a complicated mechanism of action, <sup>14</sup> and benzolamide, **BZA**, an orphan drug, <sup>15</sup> belonging to the CAI family have also been included in this study together with a large number of other aromatic and heterocyclic sulfonamide derivatives of types **1–24**.

# 2. Chemistry

Sulfonamides investigated for the inhibition of the cytosolic isozyme hCA VII, of types 1–24 are shown below. Compounds 1, 2, 4–6, 11–12, 18–20 and 23 are commercially available, whereas 3, <sup>16</sup> 7–10, <sup>17</sup> 13–17<sup>18</sup> and 21, <sup>19</sup> 22<sup>19</sup> and 24<sup>16</sup> were prepared as reported earlier by this group. The eight clinically used compounds AAZ–TPM (commercially available from Sigma–Aldrich, Merck, Alcon or Johnson & Jonson) were also assayed, since no such data are available in the literature.

#### 3. CA inhibition data

We have cloned hCA VII from a cDNA library previously described, and purified large amounts of this protein for screening studies in order to detect potent inhibitors. Similarly to the chimeric, murine protein purified earlier (mCA VII), hCA VII shows a high catalytic activity for the CO<sub>2</sub> hydration reaction, with a  $k_{\rm cat}$  of  $9.5 \times 10^5 \, {\rm s^{-1}}$  and  $k_{\rm cat}/K_{\rm m}$  of  $8.3 \times 10^7 \, {\rm M^{-1} \, s^{-1}}$  at pH 7.5 and 20 °C. The esterase activity of this isozyme is quite low (data not shown), similarly to that of mCA VII, and no detailed studies have been performed for determining the kinetic parameters of this reaction.

Inhibition data against four cytosolic CA isozymes, that is, CA I, II, VII and XIII, with a series of sulfonamides of types 1–24 and the clinically used inhibitors AAZ–TPM, for the CO<sub>2</sub> hydration reaction catalyzed by these isozymes, are shown in Table 2. Data for isozymes I, II and XIII have previously been published,<sup>2–4,8b</sup> and are presented here since they are useful when discussing the inhibition profile of the new isozyme, hCA VII. CA III inhibition data were not included, as this isozyme has a rather low affinity for sulfonamide inhibitors.<sup>2–4</sup>

16 17

The following should be noted regarding hCA VII inhibition data with the set of 32 compounds investigated here: (i) hCA VII, similarly with hCA II, is a 'sulfon-amide-avid' isozyme, being inhibited substantially ( $K_{I-S}$ )

**Table 2.** CA I, II, VII and XIII inhibition data with sulfonamides 1–24 and clinically used inhibitors. Data for hCA I and II are from Ref. 2 whereas mCA XIII data are from Ref. 8b

Inhibitor	$K_{\rm I}^{\ *} ({\rm nM})$					
	hCA I <sup>a</sup>	hCA II <sup>a</sup>	hCA VII <sup>a</sup>	mCA XIII <sup>b</sup>		
1	45,400	295	45	43		
2	25,000	240	70	32		
3	28,000	300	89	Nt		
4	78,500	320	88	Nt		
5	25,000	170	75	41		
6	21,000	160	80	43		
7	8300	60	75	50		
8	9800	110	120	56		
9	6500	40	61	54		
10	6000	70	150	50		
11	5800	63	100	Nt		
12	8400	75	210	Nt		
13	8600	60	5.2	Nt		
14	9300	19	4.3	Nt		
15	6	2	7.0	Nt		
16	164	46	5.6	2.0		
17	185	50	6.5	1.9		
18	109	33	6.8	Nt		
19	95	30	4.0	Nt		
20	690	12	5.4	Nt		
21	55	8	60	Nt		
22	21,000	125	66	Nt		
23	23,000	133	52	Nt		
24	24,000	125	68	Nt		
AAZ	250	12	2.5	17		
MZA	50	14	2.1	19		
EZA	25	8	0.78	Nt		
DCP	1200	38	26.5	23		
DZA	50,000	9	3.5	18		
BRZ	_	3	2.8	Nt		
TPM	250	5	0.87	Nt		
BZA	15	9	0.45	Nt		

Nt = not tested.

in the range of 0.45-210 nM) by all the investigated compounds 1–24 and AAZ–TPM (hCA II shows inhibition constants in the range of 2-320 nM for this set of sulfonamides/sulfamates). Thus, hCA VII is the most sensitive to these classes of CAIs among all CA isozymes investigated up to now. mCA XIII is also a sulfonamide inhibitable isozyme ( $K_I$ -s in the range of 1.9–56 nM for a series of 14 sulfonamides investigated—see Table 2) but this newly purified isozyme was available in quite limited amounts, and this is the reason why only this small number of sulfonamides has been tested in a previous work from this and Parkkila's laboratories.8 Isozyme hCA I on the other hand is much less sensitive to sulfonamide/sulfamate CAIs, showing  $K_{I}$ -s in the range of 6– 78,500 nM for the 32 derivatives included in the study; (ii) the weakest hCA VII inhibitors in this series of compounds were derivatives 8 and 10–12, which showed  $K_{I}$ -s in the range of 100–210 nM. All these derivatives, which are either halogenated sulfanilamides or benzene-1,3disulfonamide derivatives act much better as hCA II and mCA XIII than as hCA VII inhibitors, but are quite weak hCA I inhibitors; (iii) another group of derivatives, including 1-7, 9 and 21-24, are better hCA VII inhibitors as compared to the previously discussed compounds, showing inhibition constants in the range of 45– 89 nM. SAR is relatively straightforward here as all these compounds are simple benzenesulfonamide derivatives, mainly para-substituted, although several such derivatives are ortho-substituted (1 and 24) or possess two substituents (e.g., the halogenated sulfanilamides 7 and 9). It is also interesting to note that sulfonamides 1-6 and 22-24 are much better hCA VII than hCA II inhibitors, whereas in the case of 7, 9 and 21, the reverse is true (only 21 is a good hCA I inhibitor, all other discussed compounds act as rather weak inhibitors of this isozyme). On the other hand, those of these derivatives, which have been tested for the inhibition of mCA XIII showed a better activity towards this isozyme (Table 1); (iv) the clinically used compound dichlorophenamide **DCP** showed an even increased affinity for hCA VII, with an inhibition constant of 26.5 nM, rather similar to that determined against mCA XIII. DCP is on the other hand a weaker hCA II inhibitor ( $K_{\rm I}$  of 38 nM) and a much weaker hCA I inhibitor ( $K_{\rm I}$  of 1200 nM). Thus this rather simple aromatic disulfonamide has an

<sup>&</sup>lt;sup>a</sup> Human recombinant isozymes.

<sup>&</sup>lt;sup>b</sup> Murine recombinant isozyme, CO<sub>2</sub> hydrase assay method.<sup>21</sup>

<sup>\*</sup> Errors in the range of 5–10% of the shown data, from three different assays.

hCA VII inhibition profile intermediate between that of the simple aromatic compounds discussed earlier (of types 1–7, 9 and 21–24) and the more complicated derivatives belonging to the heterocyclic or polycyclic class, which will be discussed shortly; (v) very potent hCA VII inhibitory properties were observed for derivatives 13-20, AAZ, MZA, EZA, DZA, BRZ, TPM and **BZA**, which showed low nanomolar or sub-nanomolar  $K_{\rm I}$ -s for this isozyme, in the range of 0.45–7.0 nM (Table 1). Thus, among these very active hCA VII inhibitors there are some heterocyclic derivatives, belonging to the well-known ring systems present in the clinically used CAIs, of the 1,3,4-thiadiazole-2-sulfonamide type (in 13, 15, 20, AAZ and BZA), the corresponding thiadiazolines (in 14 and MZA), the benzothiazole-2sulfonamide type (in 19 and EZA) or the thienothiopyran/thiazine type of derivatives (**DZA** and **BRZ**). Other such potent hCA VII inhibitors incorporate the sulfanilyl-sulfonamide scaffold (such as 15–17) known also for a long time to induce efficient CA inhibitory activity towards many isozyme, 1,18 or belong to the sulfamate type of CAIs (i.e., topiramate TPM). Derivatives 13, 14, 16–20, as well as the clinically used compounds, are much better hCA VII than hCA II/I inhibitors, whereas 15 (aminobenzolamide) is a better hCA II than hCA VII inhibitor. Compounds 16 and 17 are better mCA XIII than hCA VII inhibitors, whereas AAZ, MZA and DZA are just the reverse, inhibiting more potently isozyme VII than isozyme XIII.

At this point, considering the very high activity as hCA VII inhibitors of a lot of derivatives (either clinically used, such as AAZ, MZA, EZA or TPM, or useful in neurophysiological experiments, such as BZA) known to possess anticonvulsant and antiepileptic activity, <sup>22,23</sup> this work strengthens the previous conclusion of Ruusuvuori et al. <sup>13</sup> that hCA VII is the main target of the antiepileptic sulfonamides/sulfamates, and allows the possibility for future drug design for obtaining compounds with a different inhibition and isozyme selectivity profiles. It should be mentioned that up to now hCA II has been considered the main isozyme responsible for the CNS effects of the sulfonamide/sulfamate inhibitors.<sup>3</sup>

## 4. Conclusions

We have cloned, purified and characterized a new CA isozyme, hCA VII, which similarly to the previously reported (chimeric) murine isozyme, shows high catalytic activity for the physiological reaction ( $k_{\rm cat}$  of  $9.5\times10^5$  s<sup>-1</sup> and  $k_{\rm cat}/K_{\rm m}$  of  $8.3\times10^7$  M<sup>-1</sup> s<sup>-1</sup> at pH 7.5 and 20 °C), and low catalytic activity as esterase. A total of 32 sulfonamides/sulfamates have been investigated as possible hCA VII inhibitors, and their inhibition profile was compared to that of the other cytosolic, catalytically active isozymes, hCA I, II and mCA XIII. hCA VII shows high affinity for all the investigated compounds, with inhibition constants in the range of 0.45–210 nM. Topiramate, ethoxzolamide, and benzolamide showed subnanomolar hCA VII inhibitory activity, whereas acetazolamide, methazolamide, dorzolamide and brin-

zolamide showed  $K_{\rm I}$ -s in the range of 2.1–3.5 nM. Dichlorophenamide was slightly less active ( $K_{\rm I}$  of 26.5 nM). A number of heterocyclic or bicyclic aromatic sulfonamides also showed excellent hCA VII inhibitory properties ( $K_{I}$ -s in the range of 4.3–7.0 nM) whereas many substituted or disubstituted benzenesulfonamides were less active ( $K_{I}$ -s in the range of 45–89 nM). The least active hCA VII inhibitors were some substituted benzene-1,3-disulfonamides as well as some halogenated sulfanilamides ( $K_{\rm I}$ -s in the range of 100–210 nM). The inhibition profile of hCA VII is rather different of that of the other cytosolic isozymes, thus providing a possibility for the design of more selective, hCA VII-specific inhibitors. These data furnish further evidence that hCA VII is the isozyme responsible for the anticonvulsant/antiepileptic activity of sulfonamides sulfamates.

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## References and notes

- Carbonic Anhydrase—Its Inhibitors and Activators; Supuran, C. T., Scozzafava, A., Conway, J., Eds.; CRC: Boca Raton (FL), USA, 2004; pp 1–373, and references cited therein.
- (a) Pastorekova, S.; Parkkila, S.; Pastorek, J.; Supuran, C. T. J. Enzym. Inhib. Med. Chem. 2004, 19, 199–229; (b) Supuran, C. T.; Vullo, D.; Manole, G.; Casini, A.; Scozzafava, A. Curr. Med. Chem.—Cardiovasc. Hematol. Agents 2004, 2, 49–68.
- (a) Supuran, C. T.; Scozzafava, A. Expert Opin. Ther. Pat. 2000, 10, 575–600;
   (b) Supuran, C. T.; Scozzafava, A. Expert Opin. Ther. Pat. 2002, 12, 217–242.
- 4. (a) Supuran, C. T.; Scozzafava, A. Curr. Med. Chem.— Imm., Endocrinol. Metab. Agents 2001, 1, 61–97; (b) Supuran, C. T.; Scozzafava, A.; Casini, A. Med. Res. Rev. 2003, 23, 146–189.
- (a) Scozzafava, A.; Mastrolorenzo, A.; Supuran, C. T. Expert Opin. Ther. Pat. 2004, 14, 667–702; (b) Supuran, C. T. Expert Opin. Ther. Pat. 2003, 13, 1545–1550.
- Hewett-Emmett, D. Evolution and Distribution of the Carbonic Anhydrase Gene Families. In *The Carbonic Anhydrases—New Horizons*; Chegwidden, W. R., Carter, N., Edwards, Y., Eds.; Birkhauser: Basel, Switzerland, 2000; pp 29–78.
- (a) Nishimori, I. Acatalytic CAs: Carbonic Anhydrase-Related Proteins CA-RPs. In Carbonic Anhydrase-Its inhibitors and Activators; Supuran, C. T., Scozzafava, A., Conway, J., Eds.; CRC: Boca Raton (FL), USA, 2004; pp 25–43; (b) Supuran, C. T. Carbonic Anhydrases: Catalytic Mechanism, Distribution and Physiological Roles. In Carbonic Anhydrase—Its Inhibitors and Activators; Supuran, C. T., Scozzafava, A., Conway, J., Eds.; CRC: Boca Raton (FL), USA, 2004; pp 1–24.
- (a) Lehtonen, J.; Shen, B.; Vihinen, M.; Casini, A.; Scozzafava, A.; Supuran, C. T.; Parkkila, A. K.; Saarnio, J.; Kivela, A. J.; Waheed, A.; Sly, W. S.; Parkkila, S. J. Biol. Chem. 2004, 279, 2719–2727; (b) Lehtonen, J.; Parkkila, S.; Vullo, D.; Casini, A.; Scozzafava, A.;

- Supuran, C. T. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 3757–3762.
- Montgomery, J. C.; Venta, P. J.; Eddy, R. L.; Fukushima, Y. S.; Shows, T. B.; Tashian, R. E. Genomics 1991, 11, 835–848.
- Lakkis, M. M.; Bergenhem, N. C.; Tashian, R. E. Biochem. Biophys. Res. Commun. 1996, 226, 268–272.
- Earnhardt, J. N.; Qian, M.; Tu, C.; Lakkis, M. M.;
   Bergenhem, N. C.; Laipis, P. J.; Tashian, R. E.; Silverman,
   D. N. *Biochemistry* 1998, 37, 10837–10845.
- 12. Lakkis, M. M.; O'Shea, K. S.; Tashian, R. E. J. Histochem. Cytochem. 1997, 45, 657–662.
- Ruusuvuori, E.; Li, H.; Huttu, K.; Palva, J. M.; Smirnov, S.; Rivera, C.; Kaila, K.; Voipio, J. *J. Neurosci.* 2004, 24, 2699–2707.
- Casini, A.; Antel, J.; Abbate, F.; Scozzafava, A.; David, S.; Waldeck, H.; Schäfer, S.; Supuran, C. T. Bioorg. Med. Chem. Lett. 2003, 13, 841–845.
- 15. Supuran, C. T.; Casini, A.; Scozzafava, A. Development of Sulfonamide Carbonic Anhydrase Inhibitors CAIs. In *Carbonic Anhydrase—Its Inhibitors and Activators*; Supuran, C. T., Scozzafava, A., Conway, J., Eds.; CRC: Boca Raton (FL), USA, 2004; pp 67–148.
- Winum, J.-Y.; Dogné, J.-M.; Casini, A.; De Leval, X.; Montero, J.-L.; Scozzafava, A.; Vullo, D.; Innocenti, A.; Supuran C. T. J. Med. Chem. 2005, in press.
- Ilies, M. A.; Vullo, D.; Pastorek, J.; Scozzafava, A.; Ilies, M.; Caproiu, M. T.; Pastorekova, S.; Supuran, C. T. *J. Med. Chem.* 2003, 46, 2187–2196.
- (a) Supuran, C. T.; İlies, M. A.; Scozzafava, A. Eur. J. Med. Chem. 1998, 33, 739–752; (b) Supuran, C. T.; Clare, B. W. Eur. J. Med. Chem. 1999, 34, 41–50; (c) Clare, B. W.; Supuran, C. T. Eur. J. Med. Chem. 1999, 34, 463–474.
- Scozzafava, A.; Menabuoni, L.; Mincione, F.; Briganti, F.; Mincione, G.; Supuran, C. T. J. Med. Chem. 1999, 42, 2641–2650.
- 20. CA VII-pGEX-2TL constructs. To produce mCA VII or hCA VII, DNA fragments covering the initiation codon and termination codon were cloned by PCR from mouse brain cDNA. The mouse sense primer was 5'-CCT GAA TTC ATG ACC GGC CAC CAC TGTT-3', which has an Eco RI site at the 5' end. The antisense primer was 5'-CCT AAG CTT TGG GGA GGT GGG AAG TTCA-3'. This primer has a Hind III site at the 5' end. The corresponding primers for human cDNA were: 5'-ACC GAA TTC ATG ACC GGC CAC CAC GGCT-3' and 5'-CCC AAG CTT

- TCA GGC CCG GAA GGA GGC-3'. PCR was performed using Sigma REDAccuTaq LA DNA polymerase mixture. The PCR mixtures were run one cycle at 98 °C for 1 min, followed by 2 cycles of 94 °C for 10 s, 55 °C for 30 s, 68 °C for 90 s, 30 cycles of 94 °C for 10 s, 65 °C for 30 s, 68 °C for 90 s, and finally 1 cycle of 68 °C for 5 min. The PCR products were cleaved with the corresponding restriction enzymes, purified and cloned into a modified pGEX-2TL vector (kindly supplied by Dr. Johan Peränen) using T4-ligase (Promega). The constructs were then transfected into *E. coli* strain BL21 for production of the CA VII protein, similarly to the procedure already described for mCA VII. 11
- 21. Khalifah, R. G. J. Biol. Chem. 1971, 246, 2561-2573, An SX.18MV-R Applied Photophysics stopped-flow instrument has been used for measuring the initial velocities by following the change in absorbance of a pH indicator. Phenol red (at a concentration of 0.2 mM) has been used as indicator, working at the absorbance maximum of 557 nm, with 10 mM Hepes (pH 7.5) as buffer, 0.1 M Na<sub>2</sub>SO<sub>4</sub> (for maintaining constant the ionic strength), following the CA-catalyzed CO2 hydration reaction for a period of 10-100 s. Saturated CO<sub>2</sub> solutions in water at 20 °C were used as substrate. The CO<sub>2</sub> concentrations ranged from 1.7 to 17 mM for the determination of the kinetic constants. For each inhibitor at least six traces of the initial 5-10% of the reaction have been used for determining the initial velocity. The uncatalyzed rates were determined in the same manner and subtracted from the total obseverd rates. The kinetic constants  $k_{\text{cat}}$  and  $k_{\text{cat}}/K_{\text{m}}$  were obtained by nonlinear least-squares methods using SigmaPlot. Stock solutions of inhibitors were prepared at a concentration of 1-3 mM (in DMSO-water 1:1, v/v) and dilutions up to 0.01 nM done with the assay buffer mentioned above. K<sub>I</sub> s of the inhibitors were determined by using Lineweaver–Burk plots, as reported earlier.  $^{15-19}$
- (a) Masereel, B.; Rolin, S.; Abbate, F.; Scozzafava, A.; Supuran, C. T. J. Med. Chem. 2002, 45, 312–320; (b) Ilies, M. A.; Masereel, B.; Rolin, S.; Scozzafava, A.; Câmpeanu, G.; Cîmpeanu, V.; Supuran, C. T. Bioorg. Med. Chem. 2004, 12, 2717–2726; (c) Chazalette, C.; Masereel, B.; Rolin, S.; Scozzafava, A.; Innocenti, A.; Supuran, C. T. Bioorg. Med. Chem. Lett. 2004, 14, 5781–5786.
- Winum, J.-Y.; Scozzafava, A.; Montero, J.-L.; Supuran,
   C. T. Expert Opin. Ther. Pat. 2004, 14, 1273–1308.